

FERMENTATION OF AGRO-BASED WASTE AND RESIDUES FROM DIFFERENT SECTORS: A REVIEW

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ABSTRACT

In India the economic importance of food and food processing sector is dominating. But the environmental issues like wastes or by-products of food industry sector is an alarming problem today. In food processing and packaging industry about one third of the food is spoilt before it reaches the consumers. This shows a huge economic loss in food sector. There are certain methods of dumping into the ground, incineration of agricultural waste and using the ash in agricultural lands. But this methods causes environmental degradation and polluting the environment. Dumping the agro-waste under normal ground causes decomposition and foul smell. Again the incineration if carried out without proper technique causes pollution. Hence, degrading the environment. So, one of the major scientific technique is carrying out different type of fermentation processes such as solid state fermentation and submerged state fermentation which could be helpful to convert this wastage into some useful high value products with the help of living cells or microorganisms in a controlled optimal atmosphere inside a specially designed suitable bioreactor or fermenter and following a suitable recovery process the final product is recovered and purified.

KEYWORDS: Fermentation, Agro-Wastes, Solid State Fermentation, Liquid State Fermentation, Substrates, Microorganisms

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INTRODUCTION

Food processing and packaging sector contributes to a huge agricultural wastage. These wastes causes different pollution problems on disposal in the ground and a huge loss of valuable biomass and nutrients is observed. Different sectors included in food industry is fruits and vegetables processing, oil seed, sugar, milk and milk products, tannery, textile and packaging industries. From these sectors during many unit operation processes a huge loss of nutrient is there in form of solid wastes like fruit peels, seeds, skin, bran, hey, whey, wood and paper pulp, fish gills, scales and liquid wastes as sugar syrups, molasses, fruit juice are wasted. (Pandey *et al.*1999). Food wastes from the food processing industry contains many nutrients in form of carbohydrates, proteins, lipids, suspended particles according to the high bio-chemical oxygen demand and chemical oxygen demand (Jayathilakan 2012) which is used by the microorganisms in the fermenter as their nutrients to grow and carry on the multiplication and fermentation process.

Concept of Fermentation

The basic concept of fermentation is to facilitate the rapid escalation, multiplication and growth of the target organisms in the nutrients provided as substrates. The desirable microorganisms metabolises the present

sugar in the substrate into chemicals like ethanol (alcohol), lactic acids. (Hui *et al.* 2004). Enzymes are protein which is synthesized as intra and extra cellular compounds. Enzymes energize and catalyze biochemical reaction with high specificity and enhance the reaction rate (Renge *et al.* 2012).

Type of Fermentation

Depending on the state of substrate the fermentation can be classified into Solid state fermentation and Liquid or submerged fermentation. In solid state fermentation the solid wastes which can be easily recycled can be directly used as substrate as high nutrient medium for growth of target microorganism. Whereas in liquid fermentation the free flowing liquid wastes from processing industries like molasses and broths are used as the nutrient medium. The solid substrates has an advantage that it can be used slowly and steadily upto a long period of time by the organisms, while the liquid substrates get easily and rapidly used up hence time to time supplementation of nutrient is required which is time consuming. While the solid state fermentation is carried out where the target organism needs less moisture content such as fungi, the liquid fermentation is suitable for high moisture activity microorganisms like bacteria. But the liquid fermentation gives advantage of easy purification and recovery after the fermentation process and is used where the product is needed in liquid form. (Subramaniam and Vimal 2012). Each type of fermentation has its own advantages.

The selection of substrate for the fermentation depends on many factors such as the nature of target organism, the nature of product to be generated. Cost and availability of the substrate also plays an important role as for high volume, low cost product the substrate should not be plenty available in low cost. Particle size should also be considered, the smaller particle size gives a good surface area for the proliferation of the target microorganisms but if the size is too small then proper enzyme action cannot occur hence resulting in poor growth of the organism resulting to incomplete fermentation and less product formation. Likewise large size particles doesn't give more surface area, though aeration and respiration is accelerated in this case. (Suganthi *et al.* 2011). The moisture content of the substrate is also required as per the target organism. When the target organism is bacteria more moisture substrate is required than in case of fungi which require less moisture to grow. The product nature determines the selection of the substrate as well as the target microorganisms.

Factors Influencing Fermentation

Factors affecting fermentation includes a number of important parameter which are needed to be taken care of during the process. Temperature, pH level, level of dissolved oxygen and dissolved carbon dioxide inside the substrate, cooling of the vessel with cold water is maintained throughout the fermentation process. The chemical composition of the substrate (whether it is in solid or liquid/submerged state), type of operation (i.e. batch, fed-batch or continuous), type of feeding inlet, type of mixer or agitator blades, power of the agitator motor, shear rate are some other factors which are the factors which should be maintained else its variation leads to different problems affecting the quality and quantity of the product by affecting its taste, smell, appearance and texture. Sometimes different toxins are created which leads to difficulty in purification.

Bioactive Compounds Extracted

Various useful components are extracted by using different suitable fermentation method according to the waste substrates available. These components are of many commercial uses and have high economic values. Many bioactive compounds such as antibiotics (Maragkoudakis *et al.*, 2009; Saykhedkar and Singhal, 2004; Ohno, 1995), pigments

(Dharmaraj et al., 2009), enzymes (Aguilar et al., 2008; Kokila and Mrudula 2010), hypercholesterolemic agents (Xie and Tang 2007; Pansuriya and Singhal, 2010), antioxidants (Tafulo et al., 2010), antihypertensive agents (Nakahara et al., 2010), antitumor agents (Ruiz-Sanchez et al., 2010), biosurfactants and bioactive peptides (Pritchard et al., 2010) have been extracted using fermentation.

Fermenter

A fermenter also known as bioreactor is generally used in a vast field of biochemical and pharmaceutical industry. A fermenter is a specially designed vessel or container made of stainless steel, glass or non-corrosive material in which residues and wastes from different agricultural sector such as peels, seeds, outer covering of fruits and vegetables, fish wastes such as gills, scales which are rich in protein, paper and wood pieces, sugar molasses and bagasses etc are converted into products of some use. The optimum temperature, pH and other suitable environment is created, maintained and controlled inside the reactor by different devices so that to encourage the growth of required specific microorganisms. All bioreactors have heterogeneous systems which deal with two or more phases like solid, liquid and gas. Sterilisation of the substrate and the bioreactor is yet another important factor which is needed to be taken care of so as to stop the growth of unwanted microorganisms other than the required ones. In a fermenter the low value waste substrates are converted and used into high value products by utilising living microorganisms or enzymes.

Fermenters are of different capacities depending on various factors. The design and capacities depends on the specific target microorganism, the optimal temperature, pH and other optimal parameters required for the growth of the target microorganisms and the quantity and quality of product. Another factor is the monetary investment and cost of running and maintenance. Bioreactors or fermenters can vary from shake conical flask (100-1000 ml), in laboratory the fermenter ranges from (1-50 L) or more, for pilot scale level (0.3- 10 m³) to plant scale (2-500 m³) depending on quality and quantity of the high value product. For products like alcoholic beverages, and lactic acids production an simple running fermenter is required without much aseptic conditions maintenance as the recovered product is of large volume and low value. For high value and low volume products such as single cell protein, antibiotics more aseptic condition is maintained and more systematic operation is required (Jagani et al.2010).The diagram of a fermenter showing different components is shown in the figure 1.

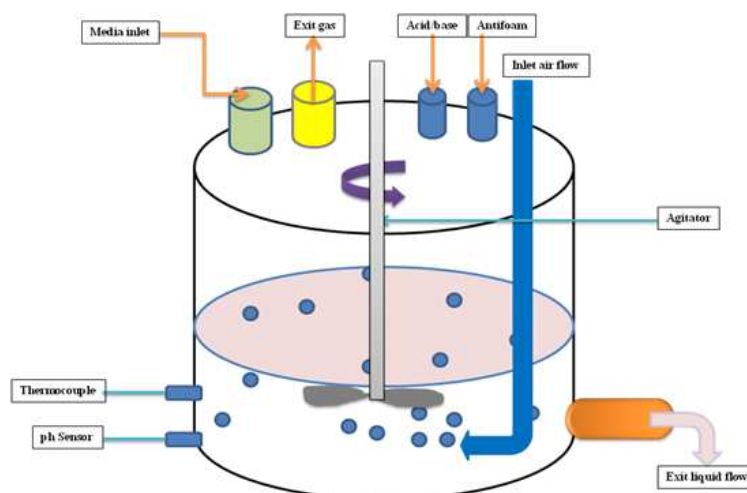


Figure 1: Diagram of a Fermenter Showing Different Basic Component

Substrates and Products from Different Agricultural Sectors Waste Products

Table 1: Different Wastes and Fermented Products Obtained

| Wastes | Microorganisms | Products | References |
|---|--|--|--|
| Substrates from Fruit and Vegetable Processing Wastes | | | |
| Wheat Bran | <i>Bacillus coagulans</i> <i>Cellumonas sp.</i> | α - amylase | Paula Monteiro de Souza <i>et al.</i> (2010) |
| Banana Fruit stalk | <i>Bacillus subtilis</i> | α - amylase | Paula Monteiro de Souza <i>et al.</i> (2010) |
| Rice Polish Wastes | <i>Penicillium notatum</i> | Glucose oxidase | S. Sabir <i>et al.</i> (2007) |
| Grape skin pulp extract, Olive oil waste effluents, Coconut milk | <i>Aureobasidium pullulans</i> | <u>Pullulans</u> (manufacture of edible films that are used in various breath freshener or oral hygiene & food additive products) | Thirumavalavan, K <i>et al.</i> (2009) |
| Fruit Pulp waste | <i>Saccharomyces cerevisiae</i> | Ethanol (With this type of fermentation, least 4% and most 8-10% alcohol production is there) | H.S.Vishwakarma <i>et al.</i> (2014) |
| Peach waste | Baker's yeast <i>Saccharomyces cerevisiae</i> | Drinkable alcohol can be obtained after application of graded distillation. Brandy which is distilled alcohol with peach aroma 6 litre brandy with 43% alcohol can be obtained from 100 kg peach waste. | Kuldip Gupta and V.K. Joshi(2000) |
| Apple pomace, | <i>Streptomyces fradiae</i> | Neomycin The highest neomycin production (2765 μ g/g substrate) was achieved with apple pomace by SSF | BM Vastrad and SE Neelagund (2011) |
| Extracted grape waste material Pressed apple pulp rich in carbon | <i>Penicillium funiculosum</i> <i>Myrothecium verrucaria</i> <i>Aspergillus niger</i> | Cellulolytic enzymes, Single cell protein | Kuzmanova, S. <i>et al.</i> (1991) |
| Dry orange Peel | <i>Aspergillus niger</i> <i>Penicillium janthinellum</i> <i>Penicillium restrictum</i> <i>Trichoderma viride</i> <i>Mucor piriformis</i> <i>Botrytis</i> <i>Ascochyta</i> <i>Absidia</i> <i>Talaromyces</i> <i>Acremonium</i> <i>Eupenicillium</i> | Citric acid (193 mg CA/g dry orange peel) | Torrado <i>et al.</i> (2011) |
| Citrus fruits peel | <i>Aerobacter</i> <i>Bacillus polymyxa</i> . <i>Xanthomonas campestris</i> | 2,3 butylene glycol (monomer) further used in synthetic rubber production Xanthan | Sterling <i>et al.</i> (1967) |
| Primarily husk, cobs, and missed kernels(maize) | <i>Aspergillus niger</i> <i>Pycnopus cinnabarinus</i> | Vanillic acid & Vanillin | Ong Khai Lun <i>et al.</i> (2014) |
| Substrates from Sugar Industry Wastes | | | |
| Molasses are source of carbon in fermentation | <i>Saccharomyces Cervicae</i> (yeast) | Ethanol L-lysine | Gasmalla <i>et al.</i> (2012) |

| | | | |
|--|--|--|---|
| processes | Corynebacterium glutamicum (for L-Lysine & L-glutamate) | L-glutamic acid. | |
| Effluents in sugar industry | <i>Candida utilis</i> | Single cell protein (SCP) | Rosma <i>et al.</i> (2006) |
| Cassava wastes | <i>Aureobasidium pullulans</i> | Exopolysachharide (Pullulans) | R.C. Ray and S.N. Moorthy (2007) |
| Cassava and sweet sorghum sugar Cane bagasse, sugar cane press Carrot-processing waste and starch | <i>Rhizopus oryzae</i> in SSF | Lactic Acid Yield of 137.0 g/l and the productivity of 1.43 g/l per hour | Soccol <i>et al.</i> (1994) |
| Cranberry pomace | <i>Lentidus edodes</i> | Ellagic acid | <ul style="list-style-type: none"> • Dhiraj A.V. and • K. Shetty (2003) |
| Production of by on untreated beet molasses in a loop airlift reactor | <i>Acetobacter xylinum</i> | Bacterial cellulose | Handbook of waste management |
| Cassava bagasse hydrolysate | <i>Rhizopus formosa</i> | Fumaric acid best producer, yielding 21.28 g/L | Handbook of waste management |
| Potato waste Cassava wastes Other starch rich vegetables | <i>Saccharomyces cerevisiae</i> | Bio-ethanol yield (87% of theoretical maximum) 29–36mL alcohol/100g of sun-dried cassava starch | Handbook of waste management |
| Potato steam peels | Photo-heterotrophic fermentation Cyano bacteria, purple bacteria | Bio-hydrogen which is a very high energy (122 kJ/g) yielding fuel | De vrije and Claassen (2003) |
| Maltodextrins result from enzymatic or acid hydrolysis of starch wastes | Filamentous fungus <i>Penicilium notatum</i> <i>Cephalosporium</i> | Penicillin, Cephalosporin, Streptomycin | Geovana <i>et al.</i> (2005) |
| Corn steep liquor is a fermented by-product of the corn wet-milling process; rich in minerals & amino acids, vitamins | <i>Saccharomyces cerevisiae</i> | Penicillin G, Amino acids, Enzymes, Bio pesticides. | Handbook of waste management |
| Substrates from Milk Industry Wastes | | | |
| Sugar is broken down, and lactic acid is produced as a by-product. | <i>Streptococcus thermophiles</i> , <i>Lactobacillus acidophilis</i> | Lactic acid fermentation (Yoghurt) in milk gives Casein | Yantyati <i>et al.</i> (2014) |
| Molasses supplemented with whey as a broth and fermentation | <i>Candida curvata</i> yeast | Microbial fat | Yantyati <i>et al.</i> (2014) |
| Substrates from Textile Sectors Wastes | | | |
| Paper, textiles, canvas and cotton wastes | <i>Myrothecium verrucaria</i> | Cellulase | Mahalaxmi M. <i>et al</i> (2011) |
| Cotton Textile Mills generate about 85,000 T of solid cellulosic wastes popularly known as Willow-dust and cyclone dust. | Methanogens Eg: <i>Methanococcus</i> , <i>Methanothrix</i> | Biogas By methane formation Raw material: 100 tonnes Total gas generation : 100 cubic m | ----- |

| Substrates from Oil Industry Wastes | | | |
|--|--|--|--|
| Plant lipids and oils cakes | <i>Penicillium</i> & other fungus by SSF | Bulk antibiotics such as: ▪ β -lactam group and (penicillins cephalosporins), ▪ Tetracyclines, ▪ Macrolides ▪ Antifungal polyenes. | A. Paithankar and A. Rewatkar (2014) |
| Oil Cakes Rapeseed palm peanut sesame soy oil cakes | <i>Pichia stipitis</i> , <i>Saccharomyces cerevisiae</i> | Bio-ethanol used for the trans-esterification of vegetable oils to produce mono ethyl esters of fatty acids as biodiesel | A. Paithankar and A. Rewatkar (2014) |
| Substrates from Tannery Wastes | | | |
| Animal fleshing, the major proteinaceous solid waste generated from leather industry | <i>Bacillus cereus</i> 1173900 by SSF | Alkaline protease Maximum protease activity of 12,310 U/g was observed at 60 h in SSF crude extract. (U) = 1 $\mu\text{mol min}^{-1}$ | Ravindran <i>et al.</i> (2011) |
| Bio- effluent | <i>Bacillus subtilis</i> | Extracellular alkaline protease | |
| Sulphite waste liquor, side product of the paper pulp manufacturing, is rich in pentose-sugars | <i>Candida yeasts</i> <i>Bacillus licheniformis</i> , <i>Bacillus stearothermophilus</i> | Ethanol α - amylase | Paula Monteiro de Souza <i>et al.</i> (2010) |
| Substrates from Fish & Fishery Wastes | | | |
| Fish Gills, Shrimp wastes | <i>Corynebacterium glutamicum</i> . | L-lysine (amino acid) | Xu J <i>et al.</i> (2014) |
| Fish wastes, mixed with 5% (w/w) sugar beet molasses | <i>Lactobacillus plantarum</i> | Fish silage excellent protein source having high biological properties for animal feeding. | Ghaly <i>et al.</i> (2013) |
| Composted fish waste | <i>Scytalidium aciabphilum</i> by SmF | Good protein yield for animal feeding. | Ghaly <i>et al.</i> (2013) |

Recovery & Purification of Fermented Products

Desired products are separated and purified after successful fermentation or enzyme reactions. Investment in recovery equipment is high. Isolation costs represent a good proportion (sometimes up to 60%) of the cost of the final product. In one antibiotic factory, recovery equipment cost four times more than the fermenter. The necessity of having a well-planned and reliable recovery process and an efficient recovery plant is therefore of utmost importance.

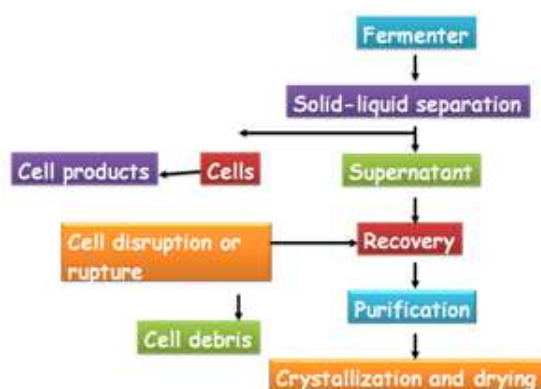


Figure 2: Flowchart for Recovery and Purification of Fermented Products

CONCLUSIONS

The food industry witness a huge loss of the products in form of peels, crushes, seeds etc during different processing and packaging processes. The food waste products are must be utilised. Fermentation is process involving microbial growths, which is widely accepted in bio chemical and pharmaceutical industry is now used in the food industry to ferment the by-products of the food industry as the substrates to carry the growth of required microorganisms into mane useful products like acids, alcohols, antibiotics etc. These products are commercially useful and add to the economy of food industry.

REFERENCES

1. Babu, K.R. and Satyanarayana, T. (1996) *Production of Bacterial Enzymes by Solid State Fermentation*. *Journal of Scientific and Industrial Research*, 55: 464-467
2. de Vrije, T. and Claassen, P.A.M. (2003) *Dark hydrogen fermentations*. In: *Bio-methane and Bio-hydrogen* (eds. J.H. Reith, R.H. Wijffels and H. Barten), pp. 150–152. *Dutch Biological Hydrogen Foundation, Smiet offset, The Hague*.
3. Dhiraj A.V. and Shetty K. (2003). *Ellagic acid production and phenolic antioxidant activity in cranberry pomace (Vaccinium macrocarpon) mediated by Lentinus edodes using a solid-state system*. *Process Biochemistry*. Vol 39, Issue 3, Pages 367–379
4. Geovana Rocha Plácido Moore' Luciana Rodrigues do Canto, Edna Regina Amante, Valdir Soldi. (2005). *Cassava and corn starch in maltodextrin production*. *Quím. Nova* vol.28 no.4 São Paulo July/Aug.
5. Ghaly AE, Ramakrishnan VV, Brooks MS, Budge SM, Dave D (2013) *Fish Processing Wastes as a Potential Source of Proteins, Amino Acids and Oils: A Critical Review*. *J Microb Biochem Technol* 5:107-129
6. Gupta K. and Joshi V.K. (2000). *Fermentative utilisation of waste from food processing industry*. *Postharvest Technology of Fruits and Vegetables: General concepts and principles*. pp 1171-1129
7. *Handbook of Food and Beverage Fermentation Technology*, edited by Y. H. Hui, Lisbeth Meunier- Goddik, Jytte Josephsen, Wai-Kit Nip, Peggy S. Stanfield
8. Hitesh Jagani*, Karteek Hebbar, Sagar S. Gang, P. Vasanth Raj, Raghu Chandrashekar H. and J.Venkata Rao. (2010). *An Overview of Fermenter and the Design Considerations to Enhance Its Productivity* *Pharmacologyonline* 1: 261-301
9. Kuzmanova, S. Vandeska, E. Dimitrovski, A. (1991). *Production of mycelial protein and cellulolytic enzymes from food wastes*. *FAO United states*
10. Mahalakshmi. M*, J. Angayarkanni, R. Rajendran, R. Rajesh (2011). *Bioconversion of cotton waste from textile mills to bioethanol by microbial saccharification and fermentation*. *Annals of Biological Research*, 2 (3) :380-388
11. Mohammed A. Gasmalla, Ruijin Y*, Mehdi N. and Su Man. (2012). *Production of Ethanol from Sudanese Sugar Cane Molasses and Evaluation of Its Quality*. *J Food Process Technol* 2012, 3:7
12. Ong Khai Lum, Tan Bee Wai and *Liew Siew Ling. (2014). *Pineapple cannery waste as a potential substrate for microbial biotransformation to produce vanillic acid and vanillin*. *International Food Research Journal* 21(3): 953-958
13. Paithankar A. and Rewatkar A. (2014). *Oil cakes as substrate for improved lipase production in solid state fermentation*. *IOSR Journal of Pharmacy and Biological Sciences*. Vol 9, Issue 4 Ver. I pp 31-38
14. Pandey, A., Selvakumar, P., Soccol, C.R., Singh - Nee Nigam, and Poonam (1999) *Solid state fermentation for the production of industrial enzymes*. *Current Science*, 77 (1): 149-162.

15. Paula Monteiro de Souza; Pérola de Oliveira e Magalhães (2010). Application of microbial α -amylase in industry – a review. *Brazilian Journal of Microbiology* (2010) 41: 850-861
16. Ravindran B., Ganesh Kumar A., Bhavani A. and Sekaran G. (2011). Solid-state fermentation for the production of alkaline protease by *Bacillus cereus* 1173900 using proteinaceous tannery solid waste. *Current science*, vol. 100, no. 5
17. Ray R.C. and Moorthy S.N. (2007). Exopolysaccharide (pullulan) production from cassava starch residue by strain *Aureobasidium pullulans* MTTC 1991. *Journal of scientific and industrial research*. Vol. 66, pp. 252-255
18. Renge V.C., Khedkar S.V. and Nandurkar N.R. (2012). Enzyme synthesis by fermentation method : a review. *Sci. Revs. Chem. Commun.*: 2(4), pp: 585-590
19. Rosma, A.* and Ooi, K. I. (2006). Production of *Candida utilis* Biomass and Intracellular Protein Content: Effect of Agitation Speed and Aeration Rate. *Malaysian Journal of Microbiology*, Vol 2(2), pp. 15-18
20. R. Suganthi, J. F. Benazir, R. Santhi, V. Ramesh Kumar, Anjana Hari, Nitya Meenakshi, K. A. Nidhiya, G. Kavitha and R. Lakshmi, (2011). Amylase Production by *Aspergillus Niger* Under Solid State Fermentation Using Agroindustrial Waste, 3(2)
21. Sabir S, Bhatti H.N.*, Zia M.A. and Sheikh M.A. (2007). Enhanced Production of Glucose Oxidase Using *Penicillium notatum* and Rice Polish. *Food Technol. Biotechnol.* 45 (4) 443–446
22. Soccol C.R., Marin B., Raimbault M. and Lebeault J.M. (1994). Potential of solid state fermentation for production of L(+)-lactic acid by *Rhizopus oryzae*. *Applied Microbiology and Biotechnology*, Volume 41, Issue 3, pp 286–290
23. Subramaniam, R. And Vimala, R. (2012). Solid state and submerged fermentation for the production of bioactive substances: a comparative study. *International journal of science and nature* vol. 3(3): 480-486
24. Sterling K. Long, Hill, and Wheaton T.A. (1967). Florida Citrus Molasses as a Fermentation Substrate. *Applied microbiology*, Sept. 1967, p. 1091-1094
25. Thirumavalavan, K, Manikkadan T.R. and Dhanasekar R. (2009). Pullulan production from coconut by-products by *Aureobasidium pullulans*. *African Journal of Biotechnology* Vol. 8 (2), pp. 254-258.
26. Torrado AM¹, Cortés S, Manuel Salgado J, Max B, Rodríguez N, Bibbins BP, Converti A, Manuel Domínguez J. (2011). Citric Acid production from orange peel wastes by solid-state fermentation. *Braz J Microbiol.* 2011 Jan; 42(1): 394-409
27. Vastrad B.M. and Neelagund S.E. (2011). Optimization and Production of Neomycin from Different Agro Industrial Wastes in Solid State Fermentation. *International Journal of Pharmaceutical Sciences and Drug Research* 2011; 3(2): 104-111
28. Vishwakarma H.S.*, Abhishek K., Singh J, Dwivedi S., Mahendra K. (2014). Production of Ethanol from Fruit Wastes by using *Saccharomyces cerevisiae*. *International Journal of Renewable Energy Technology Research*. Vol. 3, No. 10, December 2014, pp. 1-5
29. Xu J¹, Han M, Zhang J, Guo Y, Zhang W. (2014). Metabolic engineering *Corynebacterium glutamicum* for the L-lysine production by increasing the flux into L-lysine biosynthetic pathway. *Amino Acids*. Sep; 46(9): 2165-75.
30. Yantyati Widyastuti, Rohmatussolihat and Andi Febrisiantosa. (2014). The Role of Lactic Acid Bacteria in Milk Fermentation. *Food and Nutrition Sciences*, 2014, 5, 435-442